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A combined analysis of immunogenicity, antibody kinetics and vaccine efficacy from phase 2 trials of the RTS,S malaria vaccine

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Abstract

Background: The RTS,S malaria vaccine is currently undergoing phase 3 trials. High vaccine-induced antibody titres to the circumsporozoite protein (CSP) antigen have been associated with protection from infection and episodes of clinical malaria.

Methods: Using data from 5,144 participants in nine phase 2 trials, we explore predictors of vaccine immunogenicity (anti-CSP antibody titres), decay in antibody titres, and the association between antibody titres and clinical outcomes. We use empirically-observed relationships between these factors to predict vaccine efficacy in a range of scenarios.

Results: Vaccine-induced anti-CSP antibody titres were significantly associated with age (P = 0.04), adjuvant (P < 0.001), pre-vaccination anti-hepatitis B surface antigen titres (P = 0.005) and pre-vaccination anti-CSP titres (P < 0.001). Co-administration with other vaccines reduced anti-CSP antibody titres although not significantly (P = 0.095). Antibody titres showed a bi-phasic decay over time with an initial rapid decay in the first three months and a second slower decay over the next three to four years. Antibody titres were significantly associated with protection, with a titre of 51 (95% Credible Interval (CrI): 29 to 85) ELISA units/ml (EU/mL) predicted to prevent 50% of infections in children. Vaccine efficacy was predicted to decline to zero over four years in a setting with entomological inoculation rate (EIR) = 20 infectious bites per year (ibpy). Over a five-year follow-up period at an EIR = 20 ibpy, we predict RTS,S will avert 1,782 cases per 1,000 vaccinated children, 1,452 cases per 1,000 vaccinated infants, and 887 cases per 1,000 infants when co-administered with expanded programme on immunisation (EPI) vaccines. Our main study limitations include an absence of vaccine-induced cellular immune responses and short duration of follow-up in some individuals.

Conclusions: Vaccine-induced anti-CSP antibody titres and transmission intensity can explain variations in observed vaccine efficacy.

Keywords: Malaria, Vaccine, Circumsporozoite protein, Antibody, RTS,S, Phase 2 clinical trials, Mathematical model, Clinical immunity

Background

The candidate malaria vaccine RTS,S/AS01 is currently in phase 3 trials in multiple sites across sub-Saharan Africa [1,2]. Efficacy against clinical malaria over one year of follow-up was 55.8% (97.5% confidence interval (CI): 50.6% to 60.4%) in children 5- to 17-months old [1] but was significantly lower in infants 6- to 12-weeks old (31.3%, 97.5% CI: 23.6% to 38.3%) [2]. Vaccine-induced geometric mean anti-circumsporozoite protein (CSP) antibody titres following the third dose in the 6- to 12-week cohort was 209 ELISA units/ml (EU/mL) (95% CI: 197 to 222) [2] significantly lower than in the 5- to 17-month cohort (621 EU/mL, 95% CI: 592 to 652) [1]. Two potential reasons for the lower observed vaccine efficacy in the 6- to 12-week cohort have been proposed: (1) that co-administration with other expanded programme on



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immunisation (EPI) vaccines in the 6- to 12-week cohort may have interfered with the response to the RTS, S vaccine; and (2) that the 6- to 12-week cohort have reduced ability to mount a sustained and effective immune response [3], perhaps due to interference from maternal antibodies. Substantial variation in measured vaccine efficacy was also observed in the phase 2 trials, with efficacy demonstrated to depend on transmission intensity, choice of adjuvant and age at vaccination [4]. However, to date, the extent to which this variation can be explained by vaccine-induced anti-CSP antibody titres has not been explored.

Here, using data from nine phase 2 trials of the RTS,S vaccine in ten different trial sites, we investigate the association between vaccine-induced anti-CSP antibodies, their decay over time and vaccine efficacy against parasitological infection. We combine this data with a previously published model for the acquisition of clinical immunity [5] to account for reductions in efficacy against clinical malaria due to lower levels of naturally-acquired immunity in vaccinated compared to unvaccinated children. These results from heterogeneous groups in the phase 2 trials will complement the critical analyses from the multi-site phase 3 trial [1,2].

Methods

Data

Phase 2 trials of the RTS,S vaccine were identified from GlaxoSmithKline Vaccines' registry of trials and the individual-level data were provided by GSK Vaccines. Characteristics of the trials, undertaken at ten sites in six African countries, are summarised in Table 1. Healthy adults, children or infants were recruited after clinical and laboratory screening to exclude participants with clinically significant disease. Five trials considered infection as an endpoint implementing active case detection for *Plasmodium falciparum* infection (ACDi). Three trials used episodes of clinical malaria as an endpoint, one using active case detection for clinical malaria (ACDc) [6], and two using passive case detection (PCD) for clinical malaria [7,8]. One trial considered both ACDi and PCD for clinical malaria [7]. Two additional trials monitored immunogenicity but did not follow-up for clinical endpoints [9,10]. RTS,S was co-administered with other vaccines in two trials [8,11]. In total, we analysed data from 5,144 trial participants. All trials received ethical approval from relevant local ethics committees. Information on the ethical approval regarding the trials including in this analysis can be found in Additional file 1.

Immunogenicity

The method used for measuring anti-CSP antibodies was standardised and conducted in a single laboratory

[18], except for samples from The Gambia which were analysed in the Walter Reed Army Institute of Research [12]. For each participant receiving at least two doses of RTS,S/AS01 or RTS,S/AS02 we took the anti-CSP antibody titre (CSP_{peak}) measured within 21 to 30 days of the final dose to be the peak titre. Data from a fourth booster dose administered to some participants 14 months after the third dose were not included [12].

Statistical methods

We examined the effects of the following covariates on CSP_{peak} : adjuvant (AS01 *versus* AS02), age at vaccination, site-specific transmission intensity, dosing schedules (0, 1, 2 *versus* 0, 1, 7 months), number of doses received and co-administration of other vaccines. Participants were categorised according to age as follows: infants (\leq 3 months); children (>3 months and <5 years); and adults (>18 years). For each trial site, the age-corrected estimated parasite prevalence in 2- to 10-year olds in 2010 was obtained from the nearest location from the Malaria Atlas Project [17] as a proxy for transmission intensity. Trial site was included as a random effect to account for additional heterogeneity not captured by the fixed effects.

Following vaccination, the decay of antibody titres has been observed to have a short-lived phase (with titres decaying rapidly in the first few weeks), and a long-lived phase responsible for sustained vaccine-induced immunity, as has previously been observed for vaccine-induced responses to other infections [19]. To obtain estimates of anti-CSP antibody levels over time, we fitted a biphasic exponential decay model [20] to the anti-CSP antibody titres from all participants with at least two measurements. Following vaccination an individual's antibody titre CSP(t) is assumed to decay from CSP_{peak} as follows:

$$CSP(t) = CSP_{\text{peak}}\left(\rho e^{-\log(2)\frac{t}{d_s}} + (1-\rho)e^{-\log(2)\frac{t}{d_l}}\right) \qquad (1)$$

where d_s and d_l are the half-lives of the short-lived and long-lived components of the antibody response, and ρ is the proportion of the antibody response that is short-lived. Three studies included extended follow-up for longer than one year [8,14,15]. The model was fitted in a Bayesian framework using Markov Chain Monte Carlo (MCMC) methods with mixed effects used to capture between-individual variation [see Additional file 2].

We used the model-predicted anti-CSP antibody titres over time to estimate a dose–response curve for the relationship between antibody levels and protection from infection and disease using survival analysis methods [21,22]. Vaccine efficacy against infection V(t)

Table 1 Characteristics of phase 2 trial sites

Site	Participants (RTS,S)	Active vaccine	Median age (IQR)	Parasite prevalence ^a	Schedule	Peak anti-CSP titre (95% range)
Gambia [12]	250 (136)	RTS,S/AS02A	24 (19 to 34) years	70%	0,1,5,14 months	25 (13 to 43) μg/mL
Kisumu, Kenya [13]	250 (159)	RTS,S/AS02A and RTS,S/AS01B	25 (21 to 29) years	60%	0,1,2 months	34 (2 to 210) EU/mL
Manhica, Mozambique (cohort 1) [7,14]	1,589 (768)	RTS,S/AS02A	35 (24 to 48) months	40%	0,1,2 months	191 (9 to 916) EU/mL
Ilha Josina, Mozambique (cohort 2) [7,14]	411 (196)	RTS,S/AS02A	36 (24 to 45) months	45%	0,1,2 months	266 (16 to 1,390) EU/mL
Kilifi, Kenya [6,15]	447 (209)	RTS,S/AS01E	11 (8 to 14) months	35%	0,1,2 months	580 (104 to 1,922) EU/mL
Korogwe, Tanzania [6]	447 (224)	RTS,S/AS01E	12 (9 to 15) months	15%	0,1,2 months	493 (138 to 1,768) EU/mL
Kintampo, Ghana [10]	180 (180)	RTS,S/AS02D and RTS,S/AS01E	11 (8 to 14) months	80%	0,1,2 and 0,1,7 months	465 (73 to 2,632) ^b EU/mL
Kumasi, Ghana [10]	270 (270)	RTS,S/AS02D and RTS,S/AS01E	11 (7 to 13) months	35%	0,1,2 and 0,1,7 months	460 (84 to 1,785) ^b EU/mL
Lambaréné, Gabon [9]	180 (180)	RTS,S/AS02D and RTS,S/AS01E	38 (31 to 48) months	5%	0,1,2 months	198 (32 to 888) EU/mL
Bagamoyo, Tanzania [8]	209 (136)	RTS,S/AS01E	1.8 (1.7 to 1.9) months	30%	0,1,2 and 0,1,7 months ^c	167 (14 to 934) ^b EU/mL
Lambaréné, Gabon [8]	215 (139)	RTS,S/AS01E	1.5 (1.4 to 1.7) months	5%	0,1,2 and 0,1,7 months ^c	337 (97 to 1,836) ^b EU/mL
Kintampo, Ghana [8]	81 (52)	RTS,S/AS01E	1.6 (1.5 to 1.8) months	80%	0,1,2 and 0,1,7 months ^c	70 (11 to 455) ^b EU/mL
Mozambique infants [16]	214 (98)	RTS,S/AS02D	1.8 (1.8 to 2.1) months	45%	0,1,2 months	211 (6 to 1,008) EU/mL
Bagamoyo, Tanzania [11]	340 (157)	RTS,S/AS02D	1.9 (1.8 to 2) months	30%	0,1,2 months ^c	87 (1 to 572) ^b EU/mL

For participants receiving at least one dose of RTS,S the peak anti-CSP antibody titre following vaccination is presented as the median and 95% range within the cohort at each trial site. ^aAge-corrected parasite prevalence in 2- to 10-year olds taken from Malaria Atlas Project [17]; ^bindicates peak anti-CSP antibody titre in the cohort vaccinated through a 0, 1, 2 month schedule; ^cindicates co-administration with the EPI vaccines (diphtheria, tetanus, pertussis, hepatitis B, and *Haemophilus influenza* type b). CSP, circumsporozoite protein; EPI, expanded programme on immunization; EU, ELISA units; IQR, interquartile range.

is estimated from antibody titre CSP(t) according to the following dose–response curve:

$$V(t) = V_{\max}\left(1 - \frac{1}{1 + \left(\frac{CSP(t)}{\beta}\right)^{\alpha}}\right)$$
(2)

where α and β are shape parameters to be estimated, and V_{max} is the maximum efficacy against infection [21].

We assume that the entomological inoculation rate (EIR) varies between individuals to capture known heterogeneity in exposure to mosquito bites. Despite the reported seasonality in P. falciparum exposure in some of the trial sites, we make the simplifying assumption that EIR is constant over time. The rate at which an individual is exposed to malaria is then a function of (1) the EIR at the trial site and (2) their age (to account for age-dependency in biting rates). The probability that exposure results in infection is reduced by the doseresponse function for vaccine efficacy in equation (2). The probability that an infection will progress to an episode of clinical malaria will be determined by a participant's level of naturally-acquired immunity which is estimated using a previously published model [5]. Finally, the probability that a case of clinical malaria is observed is modified by a fixed effect for active or passive case detection. Parameters were estimated by fitting to the trial data in a Bayesian MCMC framework. Best fit parameters were taken to be the medians of the estimated posterior distributions. Parameters are presented with 95% credible intervals (CrI), the Bayesian analogue of confidence intervals (CI). Further details are in Additional file 2.

To assess the fit of the final model each of the phase 2 trials was re-simulated using the best fit parameters, and the results were compared to published vaccine efficacies. For each trial, the participants' peak anti-CSP antibody titre was extracted and the incidence of infection and clinical malaria simulated. Data were simulated 1,000 times, each time recording the simulated vaccine efficacy [see Additional file 2].

Estimates of efficacy and cases averted

Finally we used our fitted model to predict the expected pattern of vaccine efficacy decay against infection and clinical disease, and the cumulative number of cases averted in different transmission settings. The mean and variance of anti-CSP antibody titres following vaccination with RTS,S/AS01 by age (infants \leq 3 months; children >3 months and <5 years) and co-administration

(EPI vaccines or none) were used as an input (Table 2 and Additional file 2: Table S3). We used best fit parameters for the decay in antibody levels over time and the relationship between antibody levels and protection from infection and disease. Numbers of cases averted were estimated as the expected number of cases in an unvaccinated individual compared to an RTS,S vaccinated individual.

Results

Immunogenicity of the vaccine

Table 2 shows the impact of the tested covariates on the peak anti-CSP antibody titre following the final vaccine dose. Immunogenicity was highest among children: geometric mean anti-CSP antibody titre 465 (95% range: 41 to 5,305) EU/mL, intermediate for infants: 333 (95% range: 29 to 3,847) EU/mL and lowest for adults: 42 (95% range: 4 to 484) EU/mL. The adjuvant AS01 was significantly more immunogenic than AS02, three doses were more immunogenic than two, and RTS,S was more immunogenic when 0, 1, 2 month schedules were used rather than 0, 1, 7 month schedules. Co-administration of RTS,S with other vaccines reduced immunogenicity in infants: 82 (95% range: 7 to 941) EU/mL versus 333 (95% range: 29 to 3,847) EU/mL (P = 0.095, Table 2) although this was not statistically significant.

Higher pre-vaccination anti-CSP titres were associated with lower peak anti-CSP titres in infants (P < 0.001), but with higher peak anti-CSP titres in children (not significant, P = 0.28) and adults (P = 0.006). Higher pre-vaccination anti-hepatitis B surface antibody (HBs) titres due to prior hepatitis B vaccination were associated with higher peak anti-CSP titres in children (P = 0.005) and lower peak anti-CSP titres in adults (P = 0.002). There was no statistically significant association between peak anti-CSP titres and past exposure as measured by estimated parasite prevalence [17] (P = 0.32). Although baseline anti-CSP antibody titres are imperfect markers of past *P. falciparum* exposure, they do suggest higher levels of immunogenicity in previously exposed adults (Table 2).

Decay in vaccine-induced antibody titres

A bi-phasic exponential model for antibody decay stratified by adjuvant system was fitted to the data [see Additional file 2: Table S6]. Figure 1 shows the model fit to the data from the three studies with extended follow-up. The RTS,S/AS02 induced anti-CSP antibodies showed a lower peak following vaccination than the RTS,S/ AS01 induced antibodies (Table 2), but the pattern of antibody decay leads to similar antibody titres three years after vaccination. The bi-phasic pattern of decay in vaccine-induced antibody titres is qualitatively similar to

Covariate	Model 1 (N = 2	,659)	Model 2 (N = 1,515)		
	Estimate (95% CI)	P value	Estimate (95% CI)	P value	
Children (>3 months and <5 years)	2.59 (2.27, 2.91)	<0.001	2.50 (2.12, 2.87)	<0.001	
Infants (<3 months)	- 0.49(-0.96, -0.02)	0.04	- 0.41 (-0.91, 0.09)	0.11	
Adults (>18 years)	- 1.33 (-1.95, -0.69)	0.002	- 1.18 (-1.87, -0.48)	0.01	
log ₁₀ (CSP _{base})*children	0.05 (-0.04, 0.14)	0.28	- 0.03 (-0.14, 0.09)	0.66	
log ₁₀ (CSP _{base})*infants	- 0.58 (-0.76, -0.40)	<0.001	- 0.48 (-0.69, -0.27)	<0.001	
log ₁₀ (CSP _{base})*adults	0.24 (0.07, 0.41)	0.006	0.30 (0.11, 0.48)	0.002	
Adjuvant (AS02 versus AS01)	- 0.13 (-0.20, -0.05)	<0.001	- 0.12 (-0.19, -0.05)	<0.001	
Parasite prevalence	0.30 (-0.29, 0.89)	0.32	0.30 (-0.32, 0.93)	0.35	
Doses (2 versus 3)	- 0.46 (-0.57, -0.36)	<0.001	- 0.47 (-0.57, -0.37)	<0.001	
Schedule (017 m <i>versus</i> 012 m)	- 0.85 (-0.93, -0.77)	<0.001	- 0.84 (-0.92, -0.76)	<0.001	
Co-administration	- 0.50 (-1.04, 0.03)	0.095	- 0.52 (-1.12, 0.07)	0.13	
log ₁₀ (HBs _{base})*children	-	_	0.05 (0.02, 0.09)	0.005	
log ₁₀ (HBs _{base})*infants	-	_	- 0.04 (-0.14, 0.05)	0.34	
log ₁₀ (HBs _{base})*adults	-	-	- 0.12 (-0.20, -0.04)	0.002	

The RTS,S vaccine effect is the estimated peak anti-CSP antibody titre for a child receiving three doses of RTS,S/AS01 administered via a 0, 1, 2 month schedule without co-administration of other vaccines. The estimates presented are the regression coefficients for $log_{10}(CSP_{peak}/(EU/mL))$ against the corresponding covariates. The pre-vaccination anti-CSP and anti-HBs antibody titres are denoted CSP_{base} and HBs_{base} , respectively. Model 1 was fitted to all phase 2 trial participants with measurements of CSP_{peak} and CSP_{base} . Model 2 extends Model 1 by investigating the dependence of peak anti-CSP antibody titre on pre-vaccination anti-HBs titre, and includes all trial participants with measurements of CSP_{peak} , of CSP_{peak} . For categorical variables regression coefficients describe the association between the relevant covariate and $log_{10}(CSP_{peak})$. The regression coefficient for parasite prevalence indicates that an increase in prevalence from 0 to 1 is associated with an increase in $log_{10}(CSP_{peak})$ of 0.30. Cl, confidence interval; CSP, circumsporozoite protein; HBs, hepatitis B surface antibody.



Figure 1 Anti-CSP antibody titre dynamics. Anti-CSP antibody titre dynamics for trials with extended follow-up for longer than one year in Mozambique [14], Kilifi [23] and Ghana [10]. The black bars denote the median and 95% ranges from the data. The green and blue curves denote the antibody titres predicted by equation (1) for RTS,S/AS02 and RTS,S/AS01, respectively. Dark and light shading represents 50% and 95% of the model-predicted variation in antibody titres. We estimated that 82% (95% Crl: 80% to 83%) of the RTS,S induced antibody response was short-lived with half-life 46 (95% Crl: 43 to 49) days, the rest being long-lived with half-life 594 (95% Crl: 551 to 645) days. Crl, credible interval; CSP, circumsporozoite protein.

the decay of naturally-acquired antibody responses in African children [20].

Association between antibodies and protection from infection

Figure 2a-c shows the estimated dose–response relationships between anti-CSP antibody titre and protection for infants, children and adults. An anti-CSP antibody titre of 51 (95% CrI: 29 to 85) EU/mL was estimated to prevent 50% of infections in children and infants, and an anti-CSP antibody titre of 19 (95% CrI: 4 to 83) EU/mL was estimated to prevent 50% of infections in adults [see Additional file 2: Table S8]. Vaccine efficacy against infection and clinical malaria predicted by simulation using the estimated dose–response relationships were compared with observed efficacies in phase 2 trials (Figure 2d-e). Notably, the model captures the decline in efficacy against clinical malaria over time in a cohort of children in Kilifi, Kenya [15]. The pattern of declining efficacy in Manhica cohort 1, Mozambique [14] is also replicated, although efficacy in the first six months of follow-up is overestimated. This is possibly due to a reduction in transmission intensity in Manhica due to increased bed net coverage and other interventions not accounted for in the model [24].

Dependence of vaccine efficacy on age, exposure and time since vaccination

Figure 3a-c shows the model predicted decline in efficacy against infection over time. Efficacy decreases over time due to the decay of RTS,S induced anti-CSP antibodies (vaccine waning), with efficacy against infection in children



dropping from 54% (95% CrI: 48% to 59%) in the first year of follow-up to 27% (95% CrI: 20% to 34%) in the fifth year of follow-up. Figure 3d-f shows the predicted change in efficacy against clinical malaria over time for a range of transmission intensities. There is substantial variation in the predicted patterns of waning at different transmission intensities due to the model-predicted differential acquisition of immunity between the vaccine and control cohorts. Those in the control cohort are exposed to more infections and, hence, are predicted to develop a higher level of natural immunity over time. At EIR = 1 infectious bite per year (ibpy), the pattern of decay of efficacy against clinical malaria closely tracks the pattern observed for efficacy against infection as there is very little acquisition of natural immunity. At moderate transmission intensities (EIR = 10 to 20 ibpy), our model predicts that efficacy against clinical malaria will decay to near zero over four years. At high transmission intensities (EIR \geq 50 ibpy), efficacy against clinical malaria decays to near zero after approximately three years, after which we predict that the vaccine cohort will experience more episodes of clinical malaria than the control cohort. Figure 3g-i shows the estimated number of cumulative episodes of clinical malaria averted over time. We estimate that over a



with these estimates is presented in Additional file 2: Figure S5. EPI, expanded programme on immunization.

five-year follow-up period in a site with EIR = 20 ibpy and weekly active case detection, RTS,S averts 1,782 (95% CrI: 1,408 to 2,089) cases per 1,000 vaccinated children, 1,452 (95% CrI: 1,096 to 1,767) cases per 1,000 vaccinated infants, and 887 (95% CrI: 604 to 1,218) cases per 1,000 infants co-administered with EPI vaccines. At high transmission intensities (>20 ibpy), differences in the rate of acquisition of immunity can lead to higher incidence in the vaccine group compared to the control group at longer term, although the cumulative number of cases averted remains positive.

Discussion and conclusions

Our results demonstrate that variations in vaccine efficacy can be explained by the magnitude and duration of the RTS,S-induced anti-CSP antibody response. In turn, immunogenicity depends on age, pre-existing immunity, co-administration of other vaccinations, dosing regimen and adjuvant system. Children aged three months to five years had significantly higher vaccine-induced anti-CSP antibody titres than infants younger than three months, who in turn had higher vaccine-induced anti-CSP titres than adults. Also, the associations between pre-vaccination anti-CSP antibody titres and peak titres were negative in infants, positive in older children and non-significant in adults. Infants are thought to have less well developed capacity for immunological responses than older children [25], and our data also suggest that immunogenicity in infants is reduced further where maternal exposure to malaria was high. This might be attributable to passive immunity inhibiting the development of new immunological responses, as has been described for hepatitis B vaccination [26]. In contrast, the positive association between pre-vaccination anti-CSP titres and peak anti-CSP titres in children and adults suggests improved immunogenicity in subjects with pre-existing naturally-acquired immunity [3]. A positive association was also observed between pre-vaccination anti-HBs titres and peak anti-CSP titres in children, a result previously reported in a cohort of Gabonese children [9]. Hypothetical mechanisms underlying this association include: (1) HBs-primed B cells expressing anti-HBs antibodies capturing the RTS,S antigen and ensuring efficient presentation for CSP-specific T-cell priming; and (2) HBs-specific CD4 memory T-cells providing more rapid T-cell help to CSP-specific B cells upon stimulation by the HBs antigen in RTS,S [9].

Linear regression models without a random effect for trial site predict a statistically significant association between peak anti-CSP antibody titre and co-administration status (76% reduction, P <0.001, Additional file 2: Table S2). However, this effect may be attributable to inter-trial site variation and was not statistically significant when random effects by trial site were incorporated in the regression models (68% reduction, P = 0.095, Additional file 2: Table S1). The cohort of infants receiving RTS,S/ AS02D co-administered with EPI vaccines in Tanzania [11] had substantially lower immunogenicity than a comparable cohort of infants vaccinated with RTS,S/AS02D without co-administration in Mozambique [16]: 87 (95% range: 1 to 572) EU/mL versus 211 (95% range: 6 to 1,008) EU/mL, respectively. Statistically significant reductions in immunogenicity due to co-administration have been observed in vaccine trials of infants receiving pneumococcal [27] and Haemophilus influenza type b [28] vaccines. Hence, in context, an impact of co-administration is possible despite the lack of statistical significance.

We found anti-CSP antibody titres to be associated with protection against infection and episodes of clinical malaria [29,30], with the estimated dose–response relationship predicting increasing efficacy with increasing antibody titre across multiple trial sites (Figure 2). In particular, three of the four Prentice criteria [30] are satisfied: (1) RTS,S vaccination affects outcome (infection or clinical malaria); (2) RTS,S affects surrogates (anti-CSP antibodies); and (3) surrogates affect outcome (anti-CSP antibodies are associated with protection). The fourth criterion, that conditional upon the surrogates outcomes are independent of vaccination status, is less easily satisfied by the diverse data from the phase 2 trials. This equates to whether anti-CSP antibody titres can be used to predict vaccine efficacy. The comparison between observed and model predicted vaccine efficacies in Figure 2d,e suggests the criterion is satisfied, although there is likely to be an important role for CSP-specific CD4⁺ T cell responses which has not been accounted for. The association between anti-CSP antibody titres and protection from infection is consistent with data from other vaccine studies [31,32], as well as data from mouse models where RTS,S-induced anti-CSP antibodies have been shown to inhibit sporozoite invasion [33]. There was no evidence for a threshold anti-CSP antibody titre above which large increases in efficacy are achieved [34]. The smooth, albeit non-linear, shape of the dose-response curve is consistent with RTS,S having the profile of a leaky vaccine, but with substantial variation in efficacy.

The use of anti-CSP antibodies as the sole marker of the RTS,S-induced immune response constitutes a potential limitation of this analysis, as RTS,S-induced CSP-specific CD4⁺ T cells have been shown to be associated with protection from P. falciparum infection and episodes of clinical malaria [35]. However, analysis of immunological data from a challenge trial of RTS,S [36] suggests that anti-CSP antibody titres play a dominant role in protection from infection [21]. Furthermore, the magnitude of the RTS,S induced antibody and cell-mediated responses are correlated [36], so anti-CSP antibody titres may act as markers for cellular immunity as well as antibodymediated immunity. Efficacy against clinical malaria has also been shown to correlate with peripheral blood monocyte-to-lymphocyte ratios before vaccination [37]. Total anti-CSP immunoglobulin G (IgG) responses were measured which poses another limitation as the duration and effectiveness of the RTS,S-induced antibody responses may depend on IgG subclass [38,39].

By combining our model for the decay of anti-CSP antibody titres, the estimated relationship between antibodies and efficacy, and a model for the acquisition of immunity to clinical malaria [5], we were able to demonstrate associations between the magnitude and duration of efficacy on a number of covariates. Most striking is the dependence of efficacy against clinical malaria on estimated transmission intensity and EPI vaccine co-administration status. Increased exposure resulted in lower initial efficacy against clinical malaria in the first year of follow-up, as well as a shorter duration of protection. At longer term, the combination of decaying anti-CSP antibody titres in the vaccinated cohort and increased naturally-acquired immunity in the control cohort resulted in waning of the efficacy of RTS,S to zero or below, over a duration of follow-up dependent on transmission intensity (Figure 3). The prediction of higher incidence of clinical malaria in the vaccine

cohort compared to the control cohort during extended follow up in high transmission settings is consistent with observations of the incidence of malaria in a high exposure cohort in Kenya [15].

The variation in the characteristics of the phase 2 trials poses a potential limitation to this analysis. Differences in factors such as study populations, malaria transmission intensity, adjuvant formulation and co-administration of other vaccines complicate the analysis. For example, data on adults vaccinated with RTS,S/AS02 and followed for infection [13] are combined with data on infants vaccinated with RTS,S/AS01 and followed for clinical malaria [8]. Despite these difficulties, there is an advantage to such diversity of data as it allows systematic comparison between participants receiving different vaccination regimens at varying ages. This is in contrast to data from ongoing phase 3 trials which have more standardised vaccination regimens in more homogeneous populations [1,2]. Comparison of the varied participants in phase 2 trials with participants in phase 3 trials may provide explanations for the lower immunogenicity and efficacy observed in infants compared to children, suggesting roles for the co-administration of EPI vaccines and reductions in the immunogenicity of RTS, S-induced responses due to interference by maternallyacquired antibodies. Knowledge of the varied determinants of vaccine efficacy will allow identification of sub-populations in which RTS,S will be most effective or cost-effective [40]. Finally, the results presented in this analysis will need to be confirmed against individual level data from participants in the ongoing phase 3 trials once this becomes available [1,2].

Additional files

Additional file 1: Information on the ethical approval regarding the trials including in this analysis.

Additional file 2: Supplementary statistical methods and additional analyses.

Competing interests

The analysis for this study was conducted following a call for proposals initiated and facilitated by GSK Vaccines. Employees of GSK Vaccines reviewed and commented on early draft manuscripts, but were not involved in the final approval of the manuscript.

Authors' contributions

MTW, PB and ACG designed the study. MTW performed the analysis. JTG and JJA advised on statistical analysis. PB, AO, KP, JL, NS, SA, NO, STA, BL, KPA, SOA, EM, TA, DA, JS and JJA collected the data. MTW, PB and ACG prepared the manuscript. All authors read and approved the final manuscript.

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References

- Agnandji ST, Lell B, Soulanoudjingar SS, Fernandes JF, Abossolo BP, Conzelmann C, Methogo BG, Doucka Y, Flamen A, Mordmueller B, Issifou S, Kremsner PG, Sacarlal J, Aide P, Lanaspa M, Aponte JJ, Nhamuave A, Quelhas D, Bassat Q, Mandjate S, Macete E, Alonso P, Abdulla S, Salim N, Juma O, Shomari M, Shubis K, Machera F, Hamad AS, Minja R, *et al*: First results of phase 3 trial of RTS, S/AS01 malaria vaccine in African children. N Engl J Med 2011, 365:1863–1875.
- Mian-McCarthy S, Agnandji ST, Lell B, Fernandes JF, Abossolo BP, Methogo BG, Kabwende AL, Adegnika AA, Mordmueller B, Issifou S, Kremsner PG, Sacarlal J, Aide P, Lanaspa M, Aponte JJ, Machevo S, Acacio S, Bulo H, Sigauque B, Macete E, Alonso P, Abdulla S, Salim N, Minja R, Mpina M, Ahmed S, Ali AM, Mtoro AT, Hamad AS, RTS,S Clinical Trials Partnership, et al: A phase 3 trial of RTS, S/AS01 malaria vaccine in African infants. N Engl J Med 2012, 367:2284–2295.
- 3. Riley EM, Stewart VA: Immune mechanisms in malaria: new insights in vaccine development. *Nat Med* 2013, **19**:168–178.
- Bejon P, White MT, Olotu A, Bojang K, Lusingu J, Salim N, Otsyula N, Agnandji ST, Asante KP, Owusu-Agyei S, Abdulla S, Ghani AC: Efficacy of RTS, S malaria vaccines: individual-participant pooled analysis of phase 2 data. Lancet Infect Dis 2013, 13:319–327.
- Griffin JT, Ferguson NM, Ghani AC: Estimates of the changing age-burden of P. falciparum malaria disease in sub-Saharan Africa. *Nat Commun* 2014, 5:3136.
- Bejon P, Lusingu J, Olotu A, Leach A, Lievens M, Vekemans J, Mshamu S, Lang T, Gould J, Dubois M, Demoitié MA, Stallaert JF, Vansadia P, Carter T, Njuguna P, Awuondo KO, Malabeja A, Abdul O, Gesase S, Mturi N, Drakeley CJ, Savarese B, Villafana T, Ballou WR, Cohen J, Riley EM, Lemnge MM, Marsh K, von Seidlein L: Efficacy of RTS, S/AS01E vaccine against malaria in children 5 to 17 months of age. N Engl J Med 2008, 359:2521–2532.
- Alonso PL, Sacarlal J, Aponte JJ, Leach A, Macete E, Milman J, Mandomando I, Spiessens B, Guinovart C, Espasa M, Bassat Q, Aide P, Ofori-Anyinam O, Navia MM, Corachan S, Ceuppens M, Dubois MC, Demoitié MA, Dubovsky F, Menéndez C, Tornieporth N, Ballou WR, Thompson R, Cohen J: Efficacy of the RTS, S/AS02A vaccine against Plasmodium falciparum infection and disease in young African children: randomised controlled trial. Lancet 2004, 364:1411–1420.
- Asante KP, Abdulla S, Agnandji S, Lyimo J, Vekemans J, Soulanoudjingar S, Owusu R, Shomari M, Leach A, Jongert E, Salim N, Fernandes JF, Dosoo D, Chikawe M, Issifou S, Osei-Kwakye K, Lievens M, Paricek M, Möller T, Apanga S, Mwangoka G, Dubois MC, Madi T, Kwara E, Minja R, Hounkpatin AB, Boahen O, Kayan K, Adjei G, Chandramohan D, *et al*: Safety and efficacy of the RTS, S/AS01(E) candidate malaria vaccine given with expandedprogramme-on-immunisation vaccines: 19 month follow-up of a randomised, open-label, phase 2 trial. *Lancet Infect Dis* 2011, 11:741–749.
- 9. Lell B, Agnandji S, von Glasenapp I, Haertle S, Oyakhiromen S, Issifou S, Vekemans J, Leach A, Lievens M, Dubois MC, Demoitie MA, Carter T,

Villafana T, Ballou WR, Cohen J, Kremsner PG: A randomized trial assessing the safety and immunogenicity of AS01 and AS02 adjuvanted RTS, S malaria vaccine candidates in children in Gabon. *Plos One* 2009, 4:e7611.

- Owusu-Agyei S, Ansong D, Asante K, Owusu SK, Owusu R, Brobby NAW, Dosoo D, Akoto AO, Osei-Kwakye K, Adjei EA, Boahen KO, Sylverken J, Adjei G, Sambian D, Apanga S, Kayan K, Vekemans J, Ofori-Anyinam O, Leach A, Lievens M, Demoitie MA, Dubois MC, Cohen J, Ballou WR, Savarese B, Chandramohan D, Gyapong JO, Milligan P, Antwi S, Agbenyega T, *et al*: Randomized controlled trial of RTS, S/AS02(D) and RTS, S/AS01(E) malaria candidate vaccines given according to different schedules in Ghanaian children. *Plos One* 2009, 4:7302.
- Abdulla S, Oberholzer R, Juma O, Kubhoja S, Machera F, Membi C, Omari S, Urassa A, Mshinda H, Jumanne A, Salim N, Shomari M, Aebi T, Schellenberg DM, Carter T, Villafana T, Demoitié MA, Dubois MC, Leach A, Lievens M, Vekemans J, Cohen J, Ballou WR, Tanner M: Safety and immunogenicity of RTS, S/AS02D malaria vaccine in infants. N Engl J Med 2008, 359:2533–2544.
- Bojang KA, Milligan PJ, Pinder M, Vigneron L, Alloueche A, Kester KE, Ballou WR, Conway DJ, Reece WH, Gothard P, Yamuah L, Delchambre M, Voss G, Greenwood BM, Hill A, McAdam KP, Tornieporth N, Cohen JD, Doherty T, RTS, S Malaria Vaccine Trial Team: Efficacy of RTS, S/ASO2 malaria vaccine against Plasmodium falciparum infection in semi-immune adult men in The Gambia: a randomised trial. *Lancet* 2001, 358:1927–1934.
- Polhemus ME, Remich SA, Ogutu BR, Waitumbi JN, Otieno L, Apollo S, Cummings JF, Kester KE, Ockenhouse CF, Stewart A, Ofori-Anyinam O, Ramboer I, Cahill CP, Lievens M, Dubois MC, Demoitie MA, Leach A, Cohen J, Ballou WR, Heppner DG Jr: Evaluation of RTS, S/AS02A and RTS, S/AS01B in adults in a high malaria transmission area. *PLoS One* 2009, 4:e6465.
- Sacarlal J, Aide P, Aponte JJ, Renom M, Leach A, Mandomando I, Lievens M, Bassat Q, Lafuente S, Macete E, Vekemans J, Guinovart C, Sigaúque B, Sillman M, Milman J, Dubois MC, Demoitié MA, Thonnard J, Menéndez C, Ballou WR, Cohen J, Alonso PL: Long-term safety and efficacy of the RTS, S/AS02A malaria vaccine in Mozambican children. J Infect Dis 2009, 200:329–336.
- Olotu A, Fegan G, Wambua J, Nyangweso G, Awuondo KO, Leach A, Lievens M, Leboulleux D, Njuguna P, Peshu N, Marsh K, Bejon P: Four-year efficacy of RTS, S/AS01E and its interaction with malaria exposure. N Engl J Med 2013, 368:1111–1120.
- 16. Aponte JJ, Aide P, Renom M, Mandomando I, Bassat Q, Sacarlal J, Manaca MN, Lafuente S, Barbosa A, Leach A, Lievens M, Vekemans J, Sigauque B, Dubois MC, Demoitié MA, Sillman M, Savarese B, McNeil JG, Macete E, Ballou WR, Cohen J, Alonso PL: Safety of the RTS, S/AS02D candidate malaria vaccine in infants living in a highly endemic area of Mozambique: a double blind randomised controlled phase I/IIb trial. Lancet 2007, 370:1543–1551.
- Gething PW, Patil AP, Smith DL, Guerra CA, Elyazar IR, Johnston GL, Tatem AJ, Hay SI: A new world malaria map: Plasmodium falciparum endemicity in 2010. *Malar J* 2011, 10:378.
- Swysen C, Vekemans J, Bruls M, Oyakhirome S, Drakeley C, Kremsner P, Greenwood B, Ofori-Anyinam O, Okech B, Villafana T, Carter T, Savarese B, Duse A, Reijman A, Ingram C, Frean J, Ogutu B, Clinical Trials Partnership Committee: Development of standardized laboratory methods and quality processes for a phase III study of the RTS, S/AS01 candidate malaria vaccine. *Malar J* 2011, 10:223.
- Dalby T, Petersen JW, Harboe ZB, Krogfelt KA: Antibody responses to pertussis toxin display different kinetics after clinical Bordetella pertussis infection than after vaccination with an acellular pertussis vaccine. J Med Microbiol 2010, 59:1029–1036.
- White M, Griffin JT, Akpogheneta OJ, Conway DJ, Koram KA, Riley E, Ghani AC: Dynamics of the antibody response to *Plasmodium falciparum* infection in African children. J Infect Dis. in press.
- White MT, Bejon P, Olotu A, Griffin JT, Riley EM, Kester KE, Ockenhouse CF, Ghani AC: The relationship between RTS, S vaccine-induced antibodies, CD4(+) T cell responses and protection against plasmodium falciparum infection. *Plos One* 2013, 8:e61395.
- White MT, Griffin JT, Riley EM, Drakeley CJ, Moorman AM, Sumba PO, Kazura JW, Ghani AC, John CC: Efficacy model for antibody-mediated preerythrocytic malaria vaccines. *Proc Biol Sci* 2011, 278:1298–1305.
- Olotu A, Lusingu J, Leach A, Lievens M, Vekemans J, Msham S, Lang T, Gould J, Dubois M-C, Jongert E, Vansadia P, Carter T, Njuguna P, Awuondo KO, Malabeja A, Abdul O, Gesase S, Mturi N, Drakeley CJ, Savarese B, Villafana

T, Lapierre D, Ballou WR, Cohen J, Lemnge MM, Peshu N, Marsh K, Riley EM, von Seidlein L, Bejon P: Efficacy of RTS, S/AS01E malaria vaccine and exploratory analysis on anti-circumsporozoite antibody titres and protection in children aged 5–17 months in Kenya and Tanzania: a randomised controlled trial. *Lancet Infect Dis* 2011, **11**:102–109.

- Sharp BL, Kleinschmidt I, Streat E, Maharaj R, Barnes KI, Durrheim DN, Ridl FC, Morris N, Seocharan I, Kunene S, LA Grange JJ, Mthembu JD, Maartens F, Martin CL, Barreto A: Seven years of regional malaria control collaboration - Mozambique, South Africa, and Swaziland. *Am J Trop Med Hyg* 2007, 76:42–47.
- 25. Langhorne J, Ndungu FM, Sponaas AM, Marsh K: **Immunity to malaria:** more questions than answers. *Nat Immunol* 2008, **9**:725–732.
- Hu Y, Wu Q, Xu B, Zhou Z, Wang Z, Zhou YH: Influence of maternal antibody against hepatitis B surface antigen on active immune response to hepatitis B vaccine in infants. *Vaccine* 2008, 26:6064–6067.
- Shinefield HR, Black S, Ray P, Chang I, Lewis N, Fireman B, Hackell J, Paradiso PR, Siber G, Kohberger R, Madore DV, Malinowski FJ, Kimura A, Le C, Landaw I, Aguilar J, Hansen J: Safety and immunogenicity of heptavalent pneumococcal CRM197 conjugate vaccine in infants and toddlers. *Pediatr Infect Dis J* 1999, 18:757–763.
- Eskola J, Olander RM, Hovi T, Litmanen L, Peltola S, Kayhty H: Randomised trial of the effect of co-administration with acellular pertussis DTP vaccine on immunogenicity of Haemophilus influenzae type b conjugate vaccine. Lancet 1996, 348:1688–1692.
- Qin L, Gilbert PB, Corey L, McElrath MJ, Self SG: A framework for assessing immunological correlates of protection in vaccine trials. *J Infect Dis* 2007, 196:1304–1312.
- 30. Prentice RL: Surrogate endpoints in clinical-trials definition and operational criteria. *Stat Med* 1989, 8:431–440.
- 31. Bijker EM, Bastiaens GJ, Teirlinck AC, van Gemert GJ, Graumans W, van de Vegte-Bolmer M, Siebelink-Stoter R, Arens T, Teelen K, Nahrendorf W, Remarque EJ, Roeffen W, Jansens A, Zimmerman D, Vos M, van Schaijk BC, Wiersma J, van der Ven AJ, de Mast Q, van Lieshout L, Verweij JJ, Hermsen CC, Scholzen A, Sauerwein RW: Protection against malaria after immunization by chloroquine prophylaxis and sporozoites is mediated by preerythrocytic immunity. Proc Natl Acad Sci U S A 2013, 110:7862–7867.
- 32. Seder RA, Chang LJ, Enama ME, Zephir KL, Sarwar UN, Gordon IJ, Holman LA, James ER, Billingsley PF, Gunasekera A, Richman A, Chakravarty S, Manoj A, Velmurugan S, Li M, Ruben AJ, Li T, Eappen AG, Stafford RE, Plummer SH, Hendel CS, Novik L, Costner PJ, Mendoza FH, Saunders JG, Nason MC, Richardson JH, Murphy J, Davidson SA, Richie TL: Protection against malaria by intravenous immunization with a nonreplicating sporozoite vaccine. *Science* 2013, 341:1359–1365.
- Foquet L, Hermsen CC, van Gemert GJ, Van Braeckel E, Weening KE, Sauerwein R, Meuleman P, Leroux-Roels G: Vaccine-induced monoclonal antibodies targeting circumsporozoite protein prevent Plasmodium falciparum infection. J Clin Invest 2014, 124:140–144.
- 34. Moorthy V, Ballou WR: Immunological mechanisms underlying protection mediated by RTS, S: a review of the available data. *Malar J* 2009, 8:312.
- Olotu A, Moris PJ, Mwacharo J, Vekemans J, Kimani D, Janssens MH, Kai O, Jongert E, Lievens M, Leach A, Villafana T, Savarese B, Marsh K, Cohen J, Bejon P: Circumsporozoite-specific T cell responses in children vaccinated with RTS,S/AS01E and protection against P. falciparum clinical malaria. *PLoS One* 2011, 6:e25786.
- 36. Kester KE, Cummings JF, Ofori-Anyinam O, Ockenhouse CF, Krzych U, Moris P, Schwenk R, Nielsen RA, Debebe Z, Pinelis E, Juompan L, Williams J, Dowler M, Stewart VA, Wirtz RA, Dubois MC, Lievens M, Cohen J, Ballou WR, Heppner DG Jr, RTS, S Vaccine Evaluation Group: Randomized, double-blind, phase 2a trial of falciparum malaria vaccines RTS, S/AS01B and RTS, S/AS02A in malaria-naive adults: safety, efficacy, and immunologic associates of protection. J Infect Dis 2009, 200:337–346.
- Warimwe GM, Fletcher HA, Olotu A, Agnandji ST, Hill AV, Marsh K, Bejon P: Peripheral blood monocyte-to-lymphocyte ratio at study enrollment predicts efficacy of the RTS, S malaria vaccine: analysis of pooled phase II clinical trial data. *BMC Med* 2013, 11:184.
- Stanisic DI, Richards JS, McCallum FJ, Michon P, King CL, Schoepflin S, Gilson PR, Murphy VJ, Anders RF, Mueller I, Beeson JG: Immunoglobulin G subclass-specific responses against plasmodium falciparum merozoite antigens Are associated with control of parasitemia and protection from symptomatic illness. Infect Immun 2009, 77:1165–1174.

- Duah NO, Miles DJ, Whittle HC, Conway DJ: Acquisition of antibody isotypes against Plasmodium falciparum blood stage antigens in a birth cohort. *Parasite Immunol* 2010, 32:125–134.
- Smith T, Ross A, Maire N, Chitnis N, Studer A, Hardy D, Brooks A, Penny M, Tanner M: Ensemble modeling of the likely public health impact of a pre-erythrocytic malaria vaccine. *PloS Med* 2012, 9:e1001157.

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